



Application and advancement of microsphere as controlled delivery system: A review

Pradhi Srivastava* and Sharad Visht

Department of Pharmaceutical Technology, Meerut Institute of Engineering and Technology,
Meerut, (UP) - India

Abstract

Of late researchers have been striving hard to scale the requirements to formulate an effective dosage form to simulate controlled drug release. Microspheres have been proved to be a suitable bridge to scale this distance. Microspheres are free flowing solid particles made up of biodegradable and non-biodegradable components, ideally having a particle size less than 200 μm and can be injected by an 18 or 20 gauge needle. They can be delivered through various routes like oral, nasal, colonic, parentally, ophthalmic and transdermal etc. Various recent advancements like hollow, magnetic, microballons, floating microspheres have been contributed immensely to improve the various hindrances associated with the use of microspheres, which include site specific targeting and improved release kinetics. Current review describes the recent advancements that have been undertaken in the field of microspheres to achieve the controlled drug release, with pre-determined drug release kinetics. Author believes that the review will prove a silver lining for the aspiring researchers, thus helping them to contribute their brick in the castle entitled "microspheres".

Key-Words: Microspheres, Advancement, Applications, Preparation, Evaluation parameters

Introduction

Previously people have been using conventional dosage forms like tablet and capsule for the treatment of acute and chronic disease but these have to be taken several times to maintain the peak plasma concentration. In order to overcome this problem controlled drug delivery system was developed. The main objective of controlled drug delivery is to ensure optimum plasma drug concentration, thus enhancing efficacy and bioavailability of drug with improved patient compliance. Controlled release refers to the use of a delivery system with the objective of releasing the drug into the patient body at a predetermined rate or controlled rate, at specific times or with specific release profiles. [1]

Several problems associated with use of conventional dosage forms for the controlled release includes: The drugs which are hygroscopic in nature are not suitable for filling into capsule because moisture will be absorbed from the shell, make them brittle in nature, leading it to crumble into pieces moreover the dried solution which requires previous dilution are unsuitable for capsule because if administered as such leads to irritation into stomach.

Bioavailability problems are associated with tablets because disintegration and dissolution is required before drug is available for absorption and sometimes it also causes gastrointestinal irritation. Various advancements have been made to overcome these drawbacks of conventional dosage form and microsphere is one of them.

Microspheres are free flowing solid particles made up of biodegradable and non-biodegradable material, ideally having a particle size less than 200 μm and can be injected by an 18 or 20 gauge needle. [2] Microspheres ease sustained drug release and also reduce or eliminate gastrointestinal tract irritation. Microspheres are used to alter the drug release. Drug absorption and side effects due to irritating drugs against the gastrointestinal mucosa is improved because microspheres are made up of small particle size less than 200 μm , which are widely distributed throughout the gastrointestinal tract. [3]

Advantages [4, 5]

- Masking of odour or bitter taste.
- Improve physical stability and gastric enzyme stability.
- Better process ability (improved flowability, dispersability).
- Reduced dose size.

* Corresponding Author

E.mail: pradhisri@gmail.com

- Reduced dosing frequency therefore improves patient compliance.
- Reduced toxicity.
- Absorption window.
- Gastric irritation problem can be overcome by microsphere.
- First pass metabolism is avoided.
- Biological half-life can be enhanced.
- Improve bioavailability.
- Microsphere provides increased therapeutic efficacy and prolonged duration of action.
- Microsphere provides controlled, sustained and targeted drug delivery.
- Microsphere can be injected into body because of small size and spherical shape.

Limitation [4, 6]

- Rate of controlled release dosage form may vary due to certain factors like intrinsic and extrinsic factors.
- There is difference in the rate of release of drug from one dosage form to another dosage form.
- Low drug loading (maximum of 50%) for controlled release parental
- Dumping of dose result in failure of therapy.
- Once injected it is difficult to remove the carrier completely from the body in case of toxic effect or adverse effect.
- Parental delivery of microsphere may interact or form complexes with the blood component.

Material used for preparation microsphere [7]

Biodegradable and non-biodegradable substances are used for the preparation of microsphere. These substances involve polymers of natural and synthetic origin as well as modified natural substances.

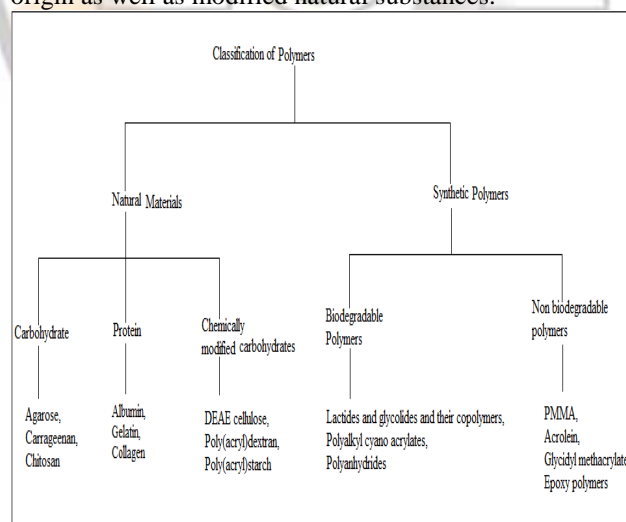


Fig. 1: Diagrammatical representation of classification of polymer

Techniques used for preparation of microspheres [5]

Microsphere can be prepared by various techniques such as:

1. Single Emulsion Technique
 - A. Heat stabilization method
 - B. Chemical stabilization method
 - C. Ionic chelation method
2. Double Emulsion Technique
3. Polymerisation Technique
 - A. Normal phase
 - a. Bulk
 - b. Suspension
 - c. Emulsion
 - B. Interfacial
4. Spray Drying Technique
5. Spray Congealing Technique
6. Solvent Extraction Technique
7. Phase Separation Co-precipitation Technique
9. Solvent Evaporation Technique

Single Emulsion Technique- [7]

Single emulsion technique is used for the preparation of microparticulate carriers for drug delivery.

Principle: Two steps involved in the preparation of microsphere by single emulsion technique are-

- Natural polymers are dissolved in aqueous solution followed by dispersion in non-aqueous solution.
- Dispersed globule is cross linked by thermal crosslinking method or by chemical crosslinkers (i.e. glutaraldehyde, formaldehyde, butanol, diacid chloride etc) or by ionic chelation method (calcium chloride).

Thermal cross linking method is not suitable for thermolabile drugs, while the disadvantage of chemical cross linking method is the excessive exposure of drug to chemicals if it is added at the time of preparation. Ionic chelation method may have demerit such as salt formation, chelation or counter ion replacement.

Double emulsion technique

Multiple emulsion or double emulsion of type w/o/w is formed during the preparation of microsphere by double emulsion technique. Natural as well as synthetic polymer are used for the preparation of microsphere by this method.

Principle: Aqueous solution of protein or polymer is dispersed in a organic lipophilic continuous phase along with the drug. Homogenization or sonication of the primary emulsion is carried out before addition to the aqueous solution of polyvinyl alcohol, which lead to the formation of multiple emulsion. Once the desired emulsion stability have been obtained, removal of solvent from the emulsion is done by solvent extraction or solvent evaporation process. Microsphere are

separated from the suspending medium by filtration, washing and drying. Incorporation of hydrophilic drug such as leutinizing hormone releasing hormone (LH-RH) agonist, vaccines, protein/peptides into microsphere can be done by Double Emulsion Technique. [7]

Example- Genistein chitosan microsphere was prepared by multiple emulsion method of type o/w/o by Wu & Li (2002). [6]
Steps involved in preparation of microsphere-

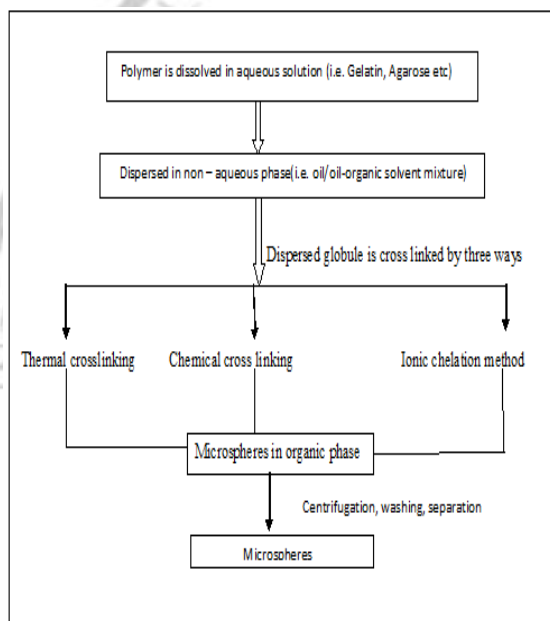


Fig. 2: Schematic representation of single emulsion based method [7]

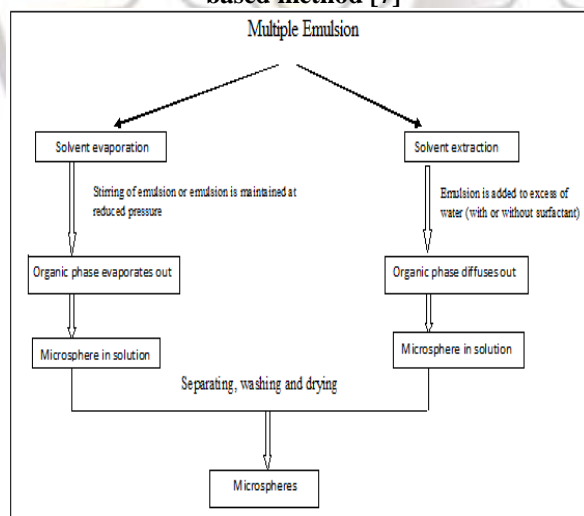


Fig. 3: Schematic representation of Microsphere preparation by Double emulsion method [7]

Polymerisation technique [9]

Microsphere can be prepared by polymerisation technique in two ways:

(A) **Normal polymerisation-** Normal polymerisation process involves various methods such as bulk polymerisation, suspension polymerisation and emulsion polymerisation for the preparation of microsphere.

(1) **Bulk polymerisation-** Bulk polymerisation process involves heating of the monomer or mixture of monomer along with the initiator and drug to initiate the polymerisation process. Polymer so obtained is moulded or fragmented as microsphere. Rate of reaction in bulk polymerisation technique can be accelerated by using the catalyst or initiator. Formation of pure polymer is the advantage of bulk polymerisation, but it cannot be used for thermolabile drug as the heat dissipated from reaction may degrade the product.

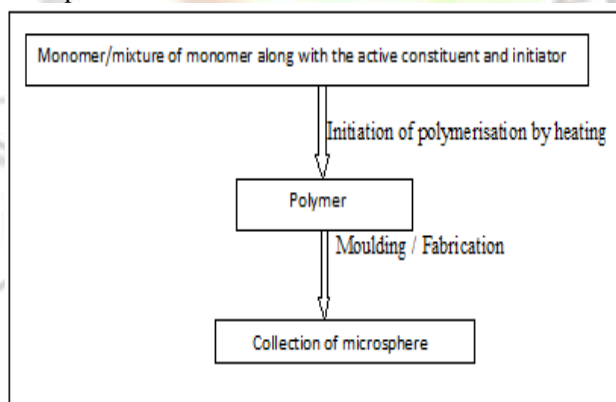


Fig. 4: Bulk polymerization method of microspheres preparation [7]

(2) **Suspension Polymerisation-** Suspension polymerisation is also known as bead polymerisation or pearl polymerisation involves heating of the monomer or a mixture of monomer along with the drug and initiator as droplet dispersion in a continuous aqueous phase. The suspension polymerisation is carried out at low temperature and heat can be easily dissipated out because continuous external phase is normally water. Polymer association with untreated monomer or other additives is the disadvantage of this technique steps used in preparation of microsphere.

(3) **Emulsion Polymerisation-** Emulsion polymerisation differs from suspension polymerisation as initiator is present in the aqueous phase, which further diffuses to the surface of the micelles or on emulsion globule.

Advantage of emulsion polymerisation with respect to bulk polymerisation is that it is used for thermolabile drug because heat can be dissipated easily and it also produces high molecular weight polymer at faster rate, but polymer association with untreated monomer or other additives is the disadvantage of this technique.

Steps used in preparation of microspheres-

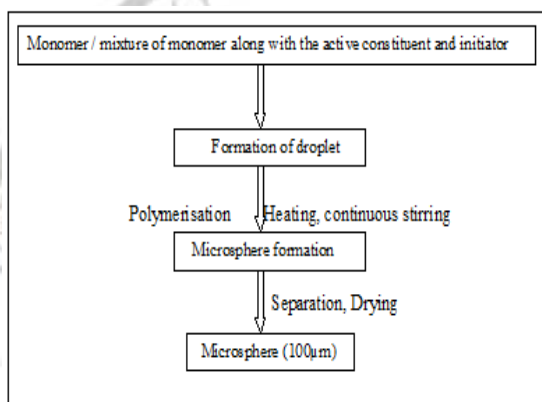


Fig. 5: Suspension polymerization method of formulation of microspheres [7]

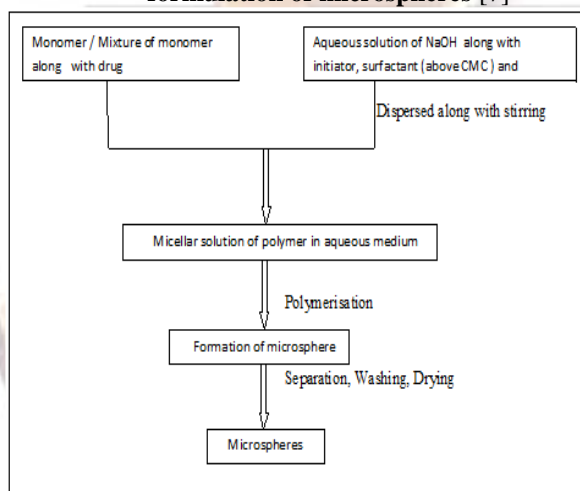


Fig. 6: Emulsion polymerization method of microspheres formulation [7]

(B) Interfacial Polymerisation– In this process, a polymer is formed between the two immiscible liquid phase by the reaction of various monomers, that envelops the dispersed phase. Continuous phase is aqueous in nature in which one monomer is dissolved and other monomer is dispersed. Monomer in either phase diffuses and polymerise at the interface rapidly, which depends upon the solubility of formed polymer in emulsion droplet. Formation of monolithic type of carrier occurs when the formed polymer is soluble in emulsion

droplet; whereas formation of capsular type of carrier (reservoir type carrier) occurs when the formed polymer is insoluble in monomer droplet. This technique is not used for the preparation of microparticles because of various drawbacks, which are as follows-

- During polymerisation degradation of drugs occurs.
- Fragility of microcapsules
- Untreated monomer causes toxicity.
- High film permeability.
- Non- biodegradability of the microparticles.

Spray Drying Technique

In Spray drying process, core material is dispersed in coating solution, in which the coating substance is dissolved and in which the polymer is insoluble, followed by atomisation of the mixture into air stream. [10]

Principle: Three steps involved during spray drying process are:

- **Atomisation**-It involves conversion of a liquid feed into fine droplet.
- **Mixing**-Mixing is carried out by passing hot air stream through spray droplets, which causes evaporation of liquid and leaving behind dried particles.
- **Dry**- Dried powder is separated from the air stream and collected. [11]

Feasibility of the process under aseptic condition is the advantage of this method; it is single stage operation, rapid and appropriate for bulk and batch manufacturing.

Disadvantage of this process is the formation of needle shape crystal when caffeine is incorporated into polymer (polylactide) by this technique which leads to incompatibility of drug and polymer (Badmeier & Chen, 1998). [12]

Example- Mucoadhesive microsphere of ondanestron for nasal administration is prepared by spray drying technique. [13]

Spray Congealing Technique- Spray congealing technique is similar as that of spray drying technique; the difference between two techniques is only that the dispersion of core material is done in a melted coating substance not in a coating solution followed by atomisation into air stream. Substance such as fatty acid, polymer, waxes, sugars which are solid at room temperature, but can be melted at certain temperature can be used in spray congealing technique. [14]

Example- Mucoadhesive microsphere can be prepared by spray congealing technique for the vaginal delivery of Econazole. [15]

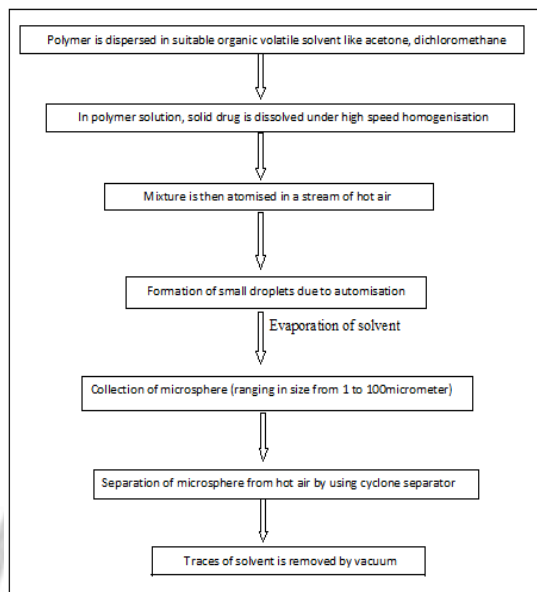


Fig. 7: Spray drying method of formation

Solvent Evaporation Technique- Solvent evaporation technique is one of the oldest and widely used methods for preparation of microsphere. When drug loading is low, this method is used for preparation of microsphere.

Example- Solvent evaporation technique is used for the preparation of microsphere of 5-Fluorouracil by using dichloromethane and acetonitrile, polyvinyl alcohol is used as processing medium to solidify the microsphere. [16] Steps involved in preparation of microsphere-

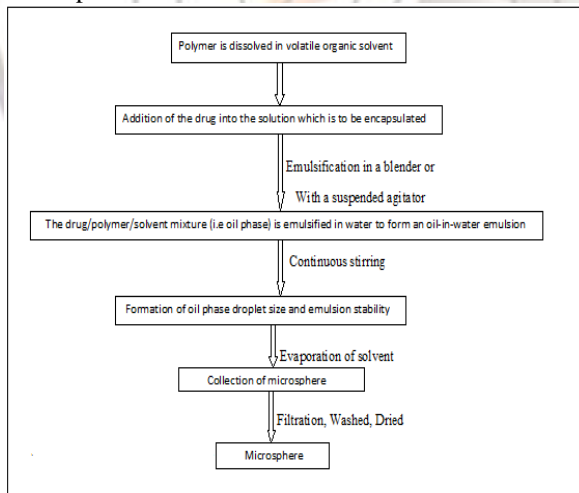


Fig. 8: Solvent evaporation technique for the microsphere formulation

Solvent Extraction Technique- In this process, removal of the organic phase is done by extraction with the organic solvent (isopropanol is used as water

miscible organic solvent). The hardening time for microsphere is decreased by removal of the organic solvent by extraction with water. Rate of solvent removal depend upon various factors such as temperature of water, ratio of emulsion volume to the water and solubility profile of the polymer. [17]

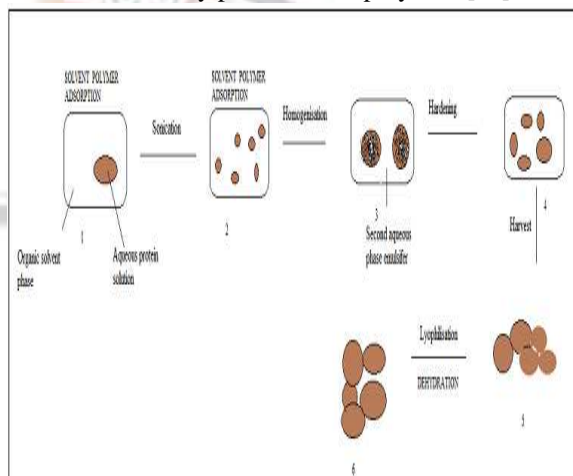


Fig. 9: Solvent extraction technique of microspheres preparation [18]

Phase Separation Coacervation Technique- In this process the solubility of polymer is decreased in the organic phase to affect the formation of the polymer rich phase known as coacervates. It is used for the preparation of reservoir type system (encapsulated water soluble drug such as proteins, peptides) and also for matrix type system (hydrophobic drug such as steroids).

Principle: Three steps involved during phase separation coacervation process are-

- Formation of three immiscible chemical phases i.e. a liquid manufacturing vehicle phase, core material phase and coating material phase.
- Deposition of the liquid coating polymer upon the core material.
- Regidization of the coating by thermal, cross linking or dissolution method.[19]

Evaluation of microsphere- Microsphere is prepared by different methods and under different conditions so that it has various microstructures which determine the release and stability of the carrier. Various parameters are evaluated for microsphere, which are as in table 2.

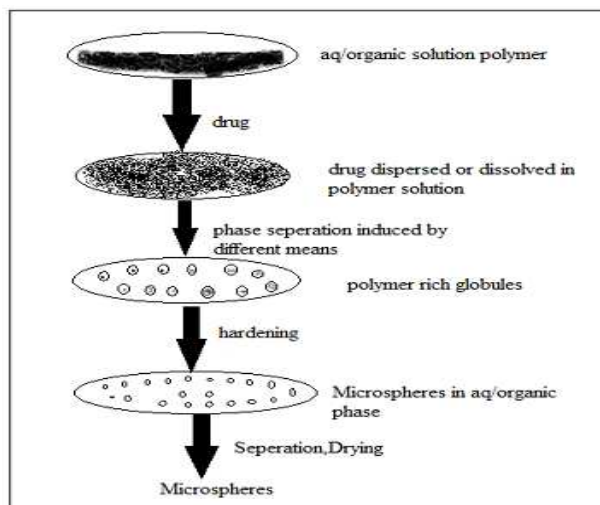


Fig. 10: Schematic representation of microsphere by phase separation coacervation technique (7)

Advancement in Microsphere

- 1. Floating microsphere-** Floating microsphere have bulk density less than gastric fluids, thus they remain floating in the stomach without affecting the gastric emptying rate. Therefore gastric retention time is increased and fluctuation in plasma drug concentration is reduced or controlled by floating microsphere. In addition it reduces dumping of dose, dosing frequency, increases therapeutic efficacy, solubility and dispersability. [34] Floating microsphere is of two types-
 - a) Effervescent type-** Swellable polymers e.g. methylcellulose, chitosan and various effervescent compound e.g. sodium bicarbonate, citric acid, tartaric acid are used for the preparation of effervescent dosage form. Floating microsphere of effervescent type liberates carbon-dioxide gas due to which the density of the system is reduced and remains in floating condition in stomach for a prolonged period of time, this result in release of drug slowly at a desired rate.
 - b) Non-effervescent type-** Highly swellable cellulose type hydrocolloids, polysaccharide and matrix forming polymer such as polycarbonate, polyacrylate are used to form non effervescent system. This is prepared by thoroughly mixing the drug and gel forming hydrocolloids. On administration, it swells up when comes in contact with gastric fluid and attain a bulk density i.e. less than 1 g/ml. [35]
- 2. Radioactive microsphere-** Radioactive microsphere is used in the same way as non-radioactive microsphere. Delivery of high

concentration of drug to the target site does not damage normal surrounding tissue. As compare to drugs, radioactivity substance is not released from the microsphere but proceed from inside a radioactive typical distance. The α emitters, β emitters and γ emitters are used as radioactive microsphere. Radioactive microsphere is used for diagnostic and therapeutic purpose. [37,38]

3. Hollow Microsphere- Hollow microspheres also called as microballoons is filled up with the drug in their outer polymer shells and are prepared by emulsion-solvent diffusion technique. The ethyl alcohol: dichloromethane solution of the drug and polymer (enteric acrylic) is transfer into a stirring aqueous solution of PVA which is thermally controlled at 40°C. Gas is generated by dispersing the polymer droplet by evaporation of dichloromethane which leads to formation of an internal cavity in microsphere of polymer with drug. The microballoons float continuously over the surface of acidic dissolution media containing surfactant for a period of more than 12 hours *in-vitro*. Different drugs can be prepared by hollow microsphere by using various materials e.g. polycarbonate, calcium alginate, Eudragit S 100 etc. [39,40]

4. Magnetic Microsphere- In magnetic microsphere small amount of magnetically targeted drug is delivering to the target site in place of high amount of free circulating drug. Those materials which response to magnetic field are used in the preparation of magnetic microsphere are chitosan, dextran etc [4]. Magnetic microsphere is used for diagnostic and therapeutic purpose. Supermagnetic iron oxides radiolabelled is used as therapeutic magnetic microsphere for imaging of liver metastasis, distinguish loop of bowel from other abdominal structure, whereas iron carbon particles radiolabelled with isotope ^{188}Re , ^{90}Y etc are used as diagnostic microsphere for liver tumours. [38]

5. Mucoadhesive Microsphere- Mucoadhesive microsphere adhere to mucus layer and release drug at desired rate. In mucoadhesive microsphere the intimate contact time with the mucus surface is increased. This results in an increased drug retention time as well as drug concentration at the targeted site [41, 42]. Mucoadhesive formulation can be administered through various routes such as nasal, gastrointestinal, buccal, Ocular etc [43]. Therapeutic efficacy is improved as well as absorption; bioavailability of normally poorly absorbed drug is improved. [44]

Conclusion

It is concluded from above that microsphere is the promising candidate for sustained and as a targeted drug delivery in GIT, liver, colon, nasal, pulmonary system, and ocular drug delivery etc. These are also used as diagnostic agent and for treatment of cancer too. Several methods of microsphere preparation give various approaches and ease to prepare various types of microspheres with different category of drugs.

References

1. Vyas S.P and Khar R.K. Controlled Drug Delivery: Concepts and Advances, Vallabh Prakashan, 1 edition 2002, Reprint 2010. Page no. 1-4.
2. Brahmankar, Biopharmaceutics and Pharmacokinetics, Vallabh Prakashan, 2 edition 2009, Page no-488.
3. V. Prasanth V, Moy Akash Chakraborty, Mathew Sam T, Mathapan Rinku. Microspheres- An Overview. International Journal of Research in Pharmaceutical and Biomedical Sciences 2011, vol.2(2):332-338.
4. Sahil Kataria, Middha Akansha, Sandhu Premjeet et al. Microsphere: A Review. International Journal of Research in Pharmacy and Chemistry 2011, 1(4):1184-1197.
5. Meena Kedar Prasad, Dangi J.S., Samal P.K. et al. Recent Advances in Microspheres Manufacturing Technology. International Journal Of Pharmacy and Technology March 2011,3:854-893.
6. Bansal Harsh, Kaur Simar Preet,Gupta A.K. et al. Microsphere : Methods Of Preparation And Applications, A Comparative Study. International Journal of Pharmaceutical Sciences Review and Research 2011, 10:69-78.
7. Vyas S.P. and Khar R.K. "Targeted and Controlled Drug Delivery: Novel Carrier System, CBS Publishers & Distributors, New Delhi ,1st edition 2002, Reprint 2011,Page no.417-451.
8. Jain N.K. "Controlled and Novel Drug Delivery" CBS Publishers & Distributors, New Delhi, 1st edition 1997, Reprint 20008, Page no.238.
9. Chaudhari Amol, Jadhav K.R., Kadam V.J. An Over View: Microsphere as A Nasal Drug Delivery System. International Journal of Pharmaceutical Sciences Review and Research 2010,5:9.
10. Bansode S.S, Banarjee S.K, Gaikwad D.D, Jadhav S.L, Thorat R.M. Microencapsulation: A Review. International Journal of Pharmaceutical Sciences Review and Research 2010,1:38-43.
11. Subrahmanyam C.V.S,Pharmaceutical Engineering ,Vallabh Prakashan,1st edition 2001,Reprint 2009, Page no.395.
12. Jain N.K. "Pharmaceutical Product Development" Publishers & Distributors, New Delhi ,1st edition 2006, Reprint 2008,Page no.127.
13. Mahajan HS. Nasal Administration of ondansteron using a novel microsphere delivery system. Pharm Dev Technol.2009; 14(2):226-32.
14. Lachman and Lieberma;The Theory and Practice Of Industrial Pharmacy, Varghese Publishing House,Bombay,1st edition1970,Reprint 1991,Page no.426.
15. Beatrice Albertini, Nadia Passerini, Marcello Di Sabatino et al. Polymer lipid based mucoadhesive prepared by spray congealing for the vaginal delivery of econazole nitrate. European journal of pharmaceutical sciences 2009, 36:591-601.
16. Behera A.L, Patil S.V, Sahoo S.K. Formulation and Characterisation of 5-Fluorouracil Microsphere by Solvent Evaporation Method, International Journal of Pharmacy and Pharmaceutical Sciences 2011, 3:32-35.
17. M Alagusundram , Madhu Sundana Chetty C., K. Umashankar . Microsphere as a Novel Drug Delivery System-A Review. International Journal of ChemTech Research 2009,1(3), :526-534.
18. Nair Rahul, Reddy B Haritha, Kumar C.K Ashok. Application of chitosan microsphere as drug carriers: A Review. Journal of Pharmaceutical Sciences & Research 2000, 1: 1-12.
19. Kumar Ankit, Sharma Pramod Kumar, Banik Arunabha. Microcapsulation as a noval drug delivery system. Internationale Pharmaceutica Scencia 2011,1:1-7.
20. Zimmer Andreas, Kreuter Jorg. Microspheres & nanoparticles used in ocular delivery systems. Advanced Drug Delivery Reviews 1995; 16:61-73.
21. Lena Pereswetoff Morath. Microspheres as novel drug delivery systems. Advanced Drug Delivery Reviews 1998; 29:185-194.
22. Rathananand Mahalaxmi, Kumar D.C, Shirwaikar A, Kumar Ravi, Kumar D.S Ampath, Prasad R.S. Preparation of mucoadhesive microspheres for nasal delivery

- by spray drying . Indian Journal of Pharmaceutical Sciences 2007; 69:651-657.
23. Singh Vandana, Chaudhary A.K. Preparation of Eudragit E100 microspheres by modified solvent evaporation method. Acta Poloniae Pharmaceutica – Drug Research 2011; 68:975-980.
 24. Madan Jyotsana, Banode Sagar, Dungi Mahesh. Mucosal Drug Delivery System. IJRAP 2010; 1:63-67.
 25. Chitosan : A Unique Pharmaceutical Excipient. Drug Delivery & Development 2005; 5. link. www.advancesearch.com.
 26. Senthil A, Sivakumar T, Narayanaswamy V.B. Mucoadhesive microspheres of oral antidiabetic drug Glipizide using different polymer. Der Pharmacia Lettere 2011; 3(2):496-506.
 27. Bittner B, Mader K, Kroll C, Barchert H-H, Kisser T. Tetracycline-HCl- loaded poly(DL-lactide-co-glycolide) microspheres prepared by a spray drying technique : influence of γ -irradiation on radical formation and polymer degradation. Journal of Controlled Release 1999; 59:23-32.
 28. Berkland Cory, Kipper Matt J, Narasimhan Balaji et al. Microsphere size, precipitation kinetics and drug distribution control drug release from biodegradable polyanhydride microspheres. Journal of Controlled Release 2004; 94:129-141.
 29. B. AppaRao, M.R. Shivalingam, Y.R. Kishore Reddy, N. Sunitha. Design and evaluation of sustained release microcapsules containing diclofenac sodium. International Journal of Pharmaceutical & Biomedicine Research 2010; 1(3):90-93.
 30. Kaza Rajesh, Pravallika.P, Reddy T.Venkata Raman, Priya K.R. Naga, Bai B.Jaya and Jyothi P.Asha. Formulation and Evaluation of Controlled Release Microspheres of Acyclovir Sodium. International Journal of Innovative Pharmaceutical Research 2011; 2(1):98-101.
 31. Narasimha Rao .R, Sampathkumar. M, Dileep Kumar.K, Vineeth P. Formulation & Evaluation of Cacicabine Microsphere. International Journal of Pharmacy & Tech 2010; 3:2598-2632.
 32. Raghavendra C. Mundargi, Namdev B. Shelke, Ajit P Rokhade et al. Formulation & in-vitro evaluation of novel starch based tableted microspheres for controlled release of ampicillin. Carbohydrate Polymer 2008; 71:42-53.
 33. Chinna Gangadhar B., Shyam Sunder R., Vimal Kumar Varma M., et al. Formulation and Evaluation of Indomethacin Microspheres using natural and synthetic polymers as Controlled Release Dosage Forms. International Journal of Drug Discovery 2010; 2:8-16.
 34. Najmuddin.M, Shelar Sachin, Ali Asgar S Patil V, Khant. Formulation & In-vitro Evaluation of floating Microsphere of Ketoprofen prepared by Emulsion Solvent Diffusion Method. International Journal of Applied Pharmaceutics 2010; 2:13-17.
 35. Nasa. Praveen, Mahat Sheefali, Sharma Deepik. Floating System A Novel Approach Towards Gastroretentive Drug Delivery System. International Journal Of Pharmacy And Pharmaceutical Sciences 2010; 2:2-7.
 36. Somwashni Sachin B, Dolas Ramdas T, Nikam Vikrant K, Gaware Vinayak M, et al. Floating multiparticulate oral sustained release drug delivery system. Journal of Chemical and Pharmaceutical Research 2011; 3(1):536-547.
 37. Hafeli Urs. Review: Radioactive Microspheres for Medical Application. Cleveland Clinic Foundation. Radiation Oncology Department T28 9500 Euclid Are., Cleveland OH 44195:1-29.
 38. Shanti. C Nithaya, Gupta Rakesh, Mahato Kumar Arun. Traditional and emerging application of microspheres:A Review. International Journal of Pharma Tech Research 2010; 2(1):675-688.
 39. Basavaraj B.V, Deveswaran R, Bharath S et al. Hollow Microspheres of Diclofenac Sodium – A Gastroretentive Controlled Delivery System. Pak. J. Pharm.Sci. 2008; 21(4):451-454.
 40. C.A Phale Sanjvani, Shinde Swapnil, Dhat Shalaka et al. Development & Evaluation of Hollow Microsphere of Clarithromycin as a Gastroretentive Drug Delivery System using Eudragit polymers. International Journal of Pharma & Biosciences. 2011; 2:344-358.
 41. Senthil. A. Mucoadhesive Microspheres. International Journal of Research in Ayurveda & Pharmacy. 2011; 2(1):55
 42. Parmar Harsad, Bakliwal Sunil, Gujarathi Nayan. Different Methods of Formulation & Evaluation of Mucoadhesive Microsphere. International Journal of Applied Biology &

- Pharmaceutical Technology. 2010; 1: 1157-1167.
43. Wang Jain, Tabata Yasuhiko, Bi Dianzhou. Evaluation of Gastric Mucoadhesive Properties of Animated Gelatin Microspheres. *Journal of Controlled Release* 2001; 73:224-231.
44. Jayvadan K Ptael, Rakesh P. Patel, Avani Famin et al. Formulation & Evaluation of Mucoadhesive Glipizide Microsphere. *AAPS PharmSciTech* 2005; 6(1): E49-E55.
45. Yellanki Shiva Kumar, Singh Jeet, Syed Jawad Ali et al. Design & Characterisation of Amoxicillin Trihydrate Mucoadhesive Microspheres for Prolonged Gastric Retention. *International Journal of Pharmaceutical Sciences & Drug Research* 2010; 2(2):112-114.
46. Trivedi P, Verma A.M.L, Garud N. Preparation & Characterisation of Aceclofenac Microsphere. *Asian Journal of Pharmaceutics* 2008; 2(2):110-115.
47. Shaji J, Puddar A, Lyer S. Brain Targeted Nasal Clonazepam Microsphere. *Indian Journal of Pharmaceutics Sciences* 2009; 71(6):1-107.

Table: 1 Tabular representation of the application of microspheres at different target sites

S.No	Target Site	Drug Use	Method Of Formulation	Applications	Polymer	Refer-ences
1	Ophthalmic	Cyclosporine, Acyclovir, Ofloxacin, Indomethacin	Cross linking, Coacervation phase separation,	Polymer hydrogels offer better acceptability. Increase precorneal drug residence time. Increase duration of efficacy of drug by using higher molecular weight polymer.	Chitosan, Alginate, Gelatin.	4,6,20.
2	Nasal	Beclomethasone, Dipropionate monohydrate, oxymetazoline hydrochloride	Single emulsion, Double emulsion method, Phase separation method	Brain achieved light concentration of drug. Improve nasal absorption. Increase bioavailability of drug.	Starch, Dextran, Albumin, Chitosan+Gelatin, DSM+LPC	3,4,21, 22.
3	Gastrointestinal drug delivery	Ranitidine hydrochloride, Melatonin, Aceclofenac, Furosemide, Amoxicillin, Prednisolone, Meloclopramid Glipizid	Solvent evaporation, By dissolving drug in polymer Cross linking	Floating and hollow microsphere can be used in drug delivery. Controlled drug release system. Decrease fluctuation in plasma drug concentration. Retention of microsphere in stomach for more than 10 hours.	Eudragit, Ethylcellulose + carbopol BSA, Gelatin	4,6,23, 24.
4	Buccal	Chlorhexide, Nefidipine, Propanolol hydrochloride.	Encapsulation, Coacervation phase separation	Prolonged release of drug in buccal cavity. Improve antimicrobial activity. Decrease in toxicity. Increase in patient compliance.	Chitosan, Sodium alginate, Gellan gum	4,6,25
5	Oral	Diazepam, Insulin	Cross linking method,	Increase patient compliance.	Chitosan, Gelatin	4,26.

			Solvent evaporation method	Multiple unit system can be produced. Avoid the exposure of high concentration of drug to mucosa. Faster drug release		
6	Intra tumoral	Fluorouracil, Cisplatin, Methotrexate, Oxantrazol.	Dry in oil, W/O in emulsion system, Combined emulsion	Delivery of therapeutically relevant concentration of drug at tumour site. Increase concentration of drug at tumour site. Decrease in side effect and toxicity of anticancer drugs. Enhance drug concentration in brain. Maximize therapeutic efficacy.	Gelatin, PLGA, Chitosan, PCL	4,6
7	Vaginal	Metronidazole, Acriflamine Methothrexate	Solvent evaporation, By dissolving drug in polymer Cross linking	Treatment of mycolic infection of genitourinary tract. Increase residence time of vaginal mucosa tissue. Adequate release and good adhesion property.	Chitosan, Gelatin, PLGA	4,6
8	Transdermal	Prednisolone, Lidocane hydrochloride, Local anesthetics	Solvent extraction method ,spray drying ,spray congealing method	Relevant packaging Sustained release action improving therapeutic efficacy Biocompatible and biodegradable polymer system can be used.	Chitosan alginate, Chitosan gelatine, PLGA	4,6,25
9	Gene delivery	Genes	Cross-linking method , polymerization method	Highly efficient and have wide range of cell targeting It also causes immune responses and oncogenic effects. Use as a carrier of DNA for gene delivery applications	Chitosan, Gelatin	4
10	Monoclonal antibodies	Amoxicillin , Ampicillin, Tetracycline, Sulfadiazine, Griseofulvine	Solvent evaporation method, Spray drying, Coacervation method	Extremely specific site targeting. Maximum stability of deliver antigen. Prolonged antigen release and lasting immunity	Chitosan, Alginate, PLGA	6,7

Table 2: Different evaluation parameters with their methods

S.No.	Evaluation Parameter	Method	Reference
1.	Particle size and shape	Light microscopy, Scanning light microscopy, Confocal scanning microscopy, Confocal fluorescence microscopy, Laser light scattering, Multisize coulter counter	27, 28
2.	Flow properties	True density, Tapped density, Bulk density, Carr's index, Hausner's ratio, Angle of repose.	29
3.	Entrapment efficiency	By allowing lysing of wash microsphere %Entrapment= Actual content/ Therotical content×100	30
4.	Density	Multivolume pycnometer	31
5.	Angle of contact	Microscope	7
6.	Isoelectric point	Microelectrophoresis	32
7.	Interaction between polymer and drug	Attenuated Total Reflectance Fourier transform Infrared Spectroscopy	7
8.	Chemical analysis	Electron Spectroscopy	31
9.	Drug release profile	USP Type Dissolution Apparatus(Rotating Paddle Type) Dialysis	3

Table 3: Drug/Polymer used in floating microsphere [36]

S.No	Drug	Polymer	Method	Result
1.	Orlistat	Eudragit S	Solvent evaporation	Increase gastric residence time of over 6 hrs.
2.	Metformin hydrochloride	Ethylcellulose	Solvent evaporation	Short processing time. High encapsulation efficiency.
3.	Cimetidine	Hdroxypropyl methylcellulose, and Ethycellulose	Solvent evaporation	Prolonged drug release (8 hrs) Remained buoyant for more than 10 hrs.
4.	Ketoprofen	Eudragit S 100 with Eudragit RL	Emulsion solvent diffusion	Drug retained in floating microparticles decreases with increased ERL content
5.	Theophylline	Mixture of cellulose acetate butyrate and Eudragit RL (1:1)		Increased residence time in stomach

Table 4: Radioactive microsphere used as diagnostic purpose

S.No	Application	Radioactive Microsphere	Reference
1.	Bone marrow imaging	^{99m} Tc sulphur colloid; ^{99m} Tc antimony sulphide colloid	37
2.	Blood flow measurement	Polystyrene microsphere labelled with γ emitters ¹⁴¹ Ce, ⁵¹ Cr	38
3.	Gated blood pool study	¹¹¹ In or ⁵¹ Cr labelled red blood cell	38
4.	Radioembolisation	⁹⁹ Tc macro aggregated human serum albumin	37
5.	Infection localisation	¹¹¹ In labelled leukocytes, ⁹⁹ Tc labelled liposomes	38

Table 5: Radioactive microsphere used as therapeutic purpose

S.No	Application	Radioactive Microsphere	Reference
1.	Local radiotherapy	⁹⁰ Y labelled polylactic acid microsphere, ¹⁶⁶ Ho acetyl acetone polylactic acid microsphere	37,38
2.	Radioembolisation of liver and spleen	⁹⁰ Y glass microsphere, ¹⁸⁸ Re Aminex A27 microsphere	38
3.	Inactivity treatment	Chromium ³² P phosphate, ⁹⁰ Y silicate	37
4.	Radiosynoviorthesis of arthritic joints	³⁵ S colloid, ¹⁸⁹ Re sulphur colloid	38
5.	Peritoneal ovarian tumour metastasis treatment, cystis brain tumour	³² P chromate	37,38

Table 6: Drug / Polymer combination used in mucoadhesive microsphere

S.No	Drug / polymer combination	Application	Reference
1.	Insulin / DSM + LPC	Help in delivery of insulin through nasal route	3,35
2.	Amoxicillin/Ethylcellulose-Carbopol	Therapeutic efficacy of drug is increased	46
3.	Aceclofenac/Eudragit	Controlled release of drug is increased	47
4.	Glipizide/Chitosan	High percentage of drug entrapment efficiency. Good mucoadhesive property	44
5.	Clonazepam/Gelatin-chitosan	High concentration of drug delivery in brain.	47